

and Phytochemistry 2020, 9(2), 1649-1653

# Journal of Pharmacognosy and Phytochemistry

Available online at www.phytojournal.com



E-188N<sub>1</sub> 2278-4136 P-ISSN: 2349-8231 Mathy tojournal.com JPP 2020; 9(2): 1649-1653 Received: 25-01-2020 Accepted: 27-02-2020

Nilesh Jawalkar Department of Zoology Dr. Babasaheb Ambedkar Marathwada University, Aurangabad, Maharashtra, India

Sureshehandra Zambare Department of Zoology Dr. Babasaheb Ambedkar Marathwada University, Anrangabad, Maharashtra, India

Amit Kulkarni Department of Microbiology, Nutan College, Schu, Maharashtra, India

# In vitro antimicrobial activity and preliminary phytochemical screening of some plant extracts

Nilesh Jawalkar, Sureshchandra Zambare and Amit Kulkarni

DOI: https://doi.org/10.22271/phyto.2020.v9.i2aa.11091

Antimicrobial activity of five solvent extracts of local plants was evaluated in vitro, with four strains of bacteria viz., Salmonella typhi, Escherichia coli, Shigella flexneri (gram -ve), Staphylococcus aureus (gram +ve) and four strains of fungi viz., Alternaria alternata, Penicillium notatum, Aspergillus niger, Penicillium digitatum microorganism. The in vitro antibacterial and antifungal activities were tested by agar disc diffusion method. The most active antibacterial plants were Vitex negundo, Tagetes erecta and antifungal plants were Xanthium strumarium, Vitex negundo and Tagetes erecta, respectively. The significant antimicrobial activities of potent extracts were compared with the standard antimicrobiotics. Ciprofloxacin and Fluconazole for bacteria as well as fungi respectively at 1 mg/ml concentration. Preliminary phytochemical analysis of V. negundo leaf, X. strumarium, M. pruriens, C. bonduc seed and T. erecta flower extracts generally revealed the presence of Alkaloids, Steroids, Terpenoids, Phenols, Saponins, Anthraquinones, Amino acids, Carotenoids, Flavonoids and Tannins at various concentrations. The results obtained in this study suggest that X. strumarium, V. negundo, T. erecta can be used in treating diseases caused by these test organisms.

Keywords: Phytochemical, antimicrobial, plant extracts, Xanthium strumarium, Vitex negundo, Tagetes erecta, ciprofloxacin, fluconazole, etc.

Bioactive compounds from vegetal sources are potential source of natural antifungic [1]. Many plants have great potential to produce new drugs which can be useful for human. Plants used in traditional medicine contain a vast array of substances that can be used to treat chronic and even infectious diseases [2]. From ancient times, plants have provided tremendous support in traditional medicine systems as well as for the development of new potential drugs in modern pharmaceutical industries. Pathogenic bacteria have always been considered as a major cause of morbidity and mortality in humans. Though the pharmaceutical companies have produced a number of new antibacterials in the last few years, resistance to these drugs has increased and it has now become a global concern [3]. Due to the increase of resistance to antibiotics, the developments of new and innovative antimicrobial agents are utmost needed. Plants have long been investigated for the potential sources of new agents. Because, they contain many bioactive secondary metabolites that can be of interest in therapeutic [4]. To consider the immense importance of medicinal plants for therapeutic target, intensive studies have been performed on different plant extracts to isolate biologically active compounds [5].

Phytochemicals are non-nutritive plant chemical that have protective or diseases preventive properties. Plant produces these chemicals to protect itself, but recent research demonstrates that many phytochemicals in leaves and seeds works differently to protect humans against

Vitex negundo (Family: Verbenaceae) commonly known as Nirgudi, is an aromatic large shrub almost found throughout India is traditionally used for the treatment of skin-ulcers, as an insecticidal, antibacterial, antifungal, for rheumatoid arthritis, gonorrhea, bronchitis, inflammation, leucoderma, enlargement of the spleen, tumors and related diseases. Scientist from different areas has studied the significant biological activities of V. negundo plant and they evaluated its antibacterial [7-12], antifungal [8, 12-13] and cytotoxic activity [6].

Xanthium strumarium L. is an annual plant belonging to the family Asteraceae generally known as common cocklebur is available generally in between October and June in India. Various parts of this plant species were found to possess useful medicinal properties such as anthelmintic [14], antifungal [15], anti-ulcerogenic [16], and anti-inflammatory [17-18] activities; it is also known to inhibit proliferation of human cancer cells in vitro [19]. [20] Srinivas et al., (2011),

orresponding Author: ilesh Jawalkar epartment of Zoology. r. Babasaheb Ambedkar arathwada University, ırangabad, Maharashtra, India

~ 1649 ~

PRINCIPAL Nutan Mahavidyalaya SELU, Dist. Parbhanl



Scanned with OKEN Scanner



forced high levels of bloactive compound group's viskaloids, phenolic acids and diterpenes and significant concentrations of saponins, glycosides, fixed oils, and phytosterols in X, stramarium, Tagetes erecta commonly known as marigold is a common omamental herbaceous shrub belonging to Asteraceae family with long history of traditional medicinal use in many countries. It is used widely in our Traditional System of Medicine for curing various diseases [21]. Since ancient time parts of this plant has been used for medicinal purposes and for the skin wash and yellow dye used as by Cherokee [12]. Marigold is commonly used in food additives as a coloring agent and as animal food in fodder (dried flower meal and extract used as supplement for poultry feed), also in tannin or dye industry [13], It is also used in medicine (folklore) and as a polson for non vertebrates and in plant pest control (24)

Coesalpinia bonduc is an Indian medicinal herb belonging to Caesalpiniaceae family, found throughout India and other tropical countries of the World [23-26]. C. bonduc seed are traditionally used in the treatment of intermittent feverasthma, colic, antiperiodic, in dyspepsia, dentrifice and Clariasis. Seed kernel is used in the treatment of orchitis, ovaritis, scrofula, useful for dispersing swellings, restraining hemorrhage in hydrocele leprosy and keeping off infectious diseases [27-29]

Mucuna pruriens (Family: Fabaceae), commonly called the velvet bean is a popular Indian medicinal plant, which is used in traditional Ayurvedic Indian medicine, for diseases including Parkinsonism and widespread in tropical and subtropical regions of the world. All parts of M. pruriens possess valuable medicinal properties and it has been investigated in various contexts, including for its anti-diabetic, aphrodisiac, anti-neoplastic, anti-epileptic, and anti-microbial activities [30] This plant is widely used in Ayurveda, which is an ancient traditional medical science that has been practiced in India since the Vedic times 1500-1000 BC [31].

## Material and Methods Collection and Identification of Plant Material

Fresh Vitex negundo leaves, Xanthium strumarium seeds, Caesalpinia bonduc seeds, Mucuna pruriens seeds and Tagetes erecta petals were collected from local, rural areas of India. Maharashtra, identification of these plants was confirmed from Department of Botany, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad. The plant material was washed under running tap water, air dried for 2-3 weeks at room temperature (37 °C - 40 °C) and pulverized in an electric grinder and stored in airtight bottles.

Preparation of the Extract 20 gm of plant powder was soaked in 200 ml ethanol, methanol, chloroform, acetone, n-Hexane solvents separately and extracts were extracted with Soxhlet apparatus.

Antimicrobial activity was planned to test using disc method. For the disc preparation Whatman filter paper No.1 was used. Whatman filter paper is folded and made a single layer. Later with the help of punching machine, it was punched into small pieces. These disc pieces were collected in the baby jar and autoclaved and preserved for further research purpose.

Proliminary Phylachemical Acrosning The major secondary metabolites like atkaloids, flavo saponins, phenols, terpenoids, anthraquinone, protein

amino acids, carbohydrates and glycosides etc. were as according to the standard procedure described by flart

Antibacterial Screening

Test organisms: A panel of bacterial strains including Escherichia coll, Salmonella typhi, Shigella flexneri (gram ve) and Staphylococcus aureus (gram +ve) were used for antibacterial activity. Standard strains were obtained from NCL (National Chemical Laboratory), Pune, India The microorganisms were maintained at 4 °C on Mueller Hinton Agar (HIMEDIA) slants.

Test organisms: A panel of fungal species including Alternaria alternata, Penicillium notatum, Aspergillus niger, Penicillium digitatum were used for antifungal activity. Standard strains obtained from Mycology laboratory of Department of Botany, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad (MS), India The microorganisms were maintained at 4 °C on Sabouraud Dextrose Agar (HIMEDIA) plates.

Preparation of the Test Solutions and Antimicrobial Assay Using Agar Disc Diffusion Methods

Antimicrobial activity was carried out according to the method of Bauer et al., (1966) [33]. Stock cultures were maintained at 4 °C on slope of Mueller Hinton Agar for bacteria and plates of Sabouraud Dextrose Agar for fungi. 200 µl of standardized cell suspensions were spread on a Mueller Hinton Agar. Extracts of all five plants (leaves and seeds) were dissolved in respective solvents and aliquot 10 µl of extracts. The samples were applied to disc paper and air dried. The air dried discs, with extract were inoculated on agar plate against selected microorganisms. After 24 hrs of incubation the results were recorded in different time interval. The effects of plant extracts were compared with Ciprofloxacin and Fluconazole as a standard for bacteria as well as fungi respectively at 1 mg/ml concentration. The results obtained by measuring diameters of the zone of inhibition in millimeter represented Average are (mm) activity, antibacterial/antifungal and + = Minimum antibacterial/antifungal activity. The data obtained was tabularized and the necessary statistical applications were

The extracts of V. negundo leaves in all five solvents showed their strong activity against E. coli. Subsequently, the extract of V. negundo leaves in three solvents, M. pruriens seeds in two solvents, T. erecta petals in three solvents showed strong activity against S. typhi as compared to different solvent extracts of X. strumarium and C. bonduc seeds. The different solvent extracts of T. erecta petals, C. bonduc seeds and V. negundo leaves showed minimum activity against S. flexneri and there is no effect of M. pruriens and X. strumarium seeds on it. The extracts of V. negundo leaves, T. erecta petals showed highest activity against bacterial strain S. aureus and other three plant species showed very less or negligible activity against it.

> PRINCIPAL Nulan Mahavidyalaya SELU, Dist. Parbhani

feds and T. erecta petals was very strong against A. negundo leaves, X. strumarium biternata. Subsequently, all the five plant extracts in different solvents showed their highest antifungal activity against P. notatum. The extracts of V. negundo leaves, C. bonduc seeds shows very less antifungal activity against A, niger and there is no any effect of X. strumarium, M. pruriens seeds and T. erecta petals in all solvent extracts against it. The antifungal activity of different solvent extracts of screened plants such as X. strumarium, C. bonduc seeds, T. erecta petals shows minimum activity as compared to V. negundo leaves against P. digitatum. There is no any effect of M. pruriens seed extracts in all the five solvents against P. digitatum. Table No. 2 summarize the microbial growth of different

solvent extracts of screened plants against gram -ve bacteria viz., E. coli, S. typhi, S. flexneri and gram +ve bacteria such as Staphylococcus aureus respectively. Table No. 3 summarize the fungicidal effect of different solvent extracts of screened

There are many differences in the antibacterial as we as antifungal effects of plant species, due to the phytochemical roperties and differences among microbes. It may quite possible that, some of the plants were ineffective in the present study do not possess antibiotic properties or the plant extracts may have contained antibacterial and antifungal constituents insufficient concentrations. The drying process may have caused conformational changes to occur in some of the chemical constituents found in these plants species.

The secondary metabolites like tannins, flavonoids, steroids, saponins, alkaloids phenols, terpenoids, anthraquinone, proteins and amino acids present in trace amount in some of the plants are showed in Table No. 1.

### Discussion

Each of the extract tested in the present study displayed antibacterial as well as antifungal activity on at least 2-4 bacterial and 2-4 fungal strains tested. However, differences were observed between antibacterial and antifungal activities of the extracts. These differences may be due to change in the chemical composition of these extracts as the bioactive compounds from plants have many effects including antibacterial and antiviral properties [34-35].

Rose and Cathrine (2011) 1161, reported E coli and Streptococcus mutans were resistant to three different solven extracts of V. negundo leaves viz., petroleum ethe dichloromethane and ethanol. They also reported the Klehstela pneumoniae, Vihrio cholerae, Streptococci mutans, Eschericia coli have inhibitory activity next only the Salmonella paratyphi. Lolge et al., (2016) [37], in three C. auriculata, A. mexicana, S. trifoliatus plant extracts in four different solvents; acetone, methanol, ethanol and chloroform showed their antibacterial activity against E, coll and S, aureus, Islam et al., (2013) [18], studied; Leaf extract of V. negundo in methanol at 10 mg/ml exhibited a variable growth inhibition capacity against all bacterial species viz., Bacillus subtilis, Staphylococcus aureus, Pseudomonas aeruginosa, Escherichia coli and Salmonella typhi. Devkota and Das (2015) [39], showed the methanolic extract of X. strumarium shows zone of inhibition with K. pneumoniae, E. faecalis, B. subtilis, P .mirabilis, S. aureus but it was ineffective against E. coli. Similarly, distilled water extract of X. strumarium showed zone of inhibition with E. faecalis, B. subtilis, P. mirabilis, S. aureus and it was incapable with E. coli and K. pneumonia. The leaf extract of X. strumarium in aqueous, ethanol and n-hexane solvents, inhibited S. aureus and not E. coli, K. pneumoniae and B. subtilis [40], but in the present study only n-Hexane extract of X. strumarium showed antibacterial effects against S. aureus. The extract of chloroform of X. strumarium inhibited S. aureus, B. subtilis, E. coli and ethyl acetate extract inhibited S. aureus [41], but no inhibition of any bacterial growth was observed by aqueous extract however we haven't found any activity of chloroform extract of X. strumarium seed against S. aureus and E. coli. These differences in result may be due to the difference in time of collection of plant material, difference of solvent, dose and habitat of plant material. Verma and Verma (2012) [21], studied the antibacterial potential of ethanolic extracts from leaves, flower, stem and root of plant T. erecta and reported that S. lutea, B. circulence, B. subtilis were most sensitive to the leaf extract while E. coli and S. aureus were least sensitive to the ethanolic extract. Flower extract showed significant antibacterial activity against S. lutea, E. coli, B. circulence.

Table 1: Preliminary qualitative phytochemical analysis of five screened plants species

l	DI 4	Tanning	Saponins	Phenols	Flavonoids	Alkaloids	Steroids	Terpenoids	Anthraquinone	Amino acid
Plant species	Plant part	Tannins	Saponins	1 inclinois	•		-		+	+
Visar magninda	Leaves	+	+	+	+	+				+
Vitex negundo		-	- 1	+	+	+	-	+		-
Xanthium strumarium	Seed	+	т —	-			+	+		-
Caesalpinia bonduc		+	+	-	+			+	-	+
	Seed	+	+	+		+				+
Mucuna pruriens Tagetes erecta	Petals	+	+	+	+	+	+		- Lostoni	

Table 2: Antibacterial activity of different solvent extracts of screened plants against Gram -ve and Gram +ve bacteria.

Table 2: Antibacte	mai activity		- (Ci - Covenin)	Ethanol	Methanol	Chloroform	Acetone	n-Hexane						
- Diant Species	Plant Part		Control (Ciprolloxacili)	+++	+++	++	+++	+						
o. Plant Species		E. coli (Gram -ve)				+	++	-						
1. Vitex negundo	Leaves	S typhi (Gram -ve)	+++				++	++						
		S. Ogrneri (Gram -ve)	+++			++	+++	-						
The neg		C. gurgus (Gram +ve)	+++	+++	+++		-	-						
		5. aureus (Gram -ve)	+++	-			+	+						
	Seed S	E. Coll (Gram ve)	+++	-	+	-	-	-						
		S. typhi (Grain -ve)	+++	-		-	-	+						
Xanthium strumarium		S. flexneri (Gram -ve)		-	-	-	-							
		S. aureus (Gram +ve)			-	-	-	-						
									E. coli (Gram -ve)		+	+	++	+
	esalpinia Seed S. typhi	S typhi (Gram -ve)			+	++	+	-						
Caesalpinia		florneri (Gram -ve)	+++			+	+	-						
bonduc		(Gram +ve)	+++	+										
		aureus (Grain 117)	~ 1651 ~					4						
	Vitex negundo	Vitex negundo  Leaves  Xanthium strumarium  Caesalpinia  Seed	Vitex negundo  Leaves    Leaves   E. coli (Gram -ve)	Vitex negundo   Leaves	Vitex negundo         Plant Part         Bacterium         Control (Ciprofloxacin) Ethanol           Vitex negundo         Leaves              E. coli (Gram -ve)	Vitex negundo         Leaves         E. coli (Gram -ve)         +++  <	O.         Plant Species         Plant Part         Bacterium         Control (Ciprofloxacin) Ethanol Methanol Chrostors and the product of th	Vitex negundo         Plant Part         Bacterium         Control (Ciprofloxacin) Ethanol Methanol Chrostors and Ethanol Methanol Methanol Methanol Chrostors and Ethanol Methanol Methanol Chrostors and Ethanol Chrostors and						

PRINCIPAL Nutan Mahavidyalaya SELU, Dist. Parbhani



Scanned with OKEN Scanner



S	
Ξ.	
òi	
ш	
$\exists$	
=	
_	
æ	
Ô	
_	
<	
<	
_:	
ᇁ	
_	
_	
$\circ$	
-	
$\sim$	
ш	
-	
_	
S	
C	
Ò	
=	
_	
_	
<u></u>	
m	

1	-		1				http.	henry ch	ytologimale	ţi
1	Mucuna pruriens	Seed	E. coli (Gram -ve) S. typhi (Gram -ve) S. flexneri (Gram -ve)	+++	-:	-:-		<u>:</u>		
5.	Tagetes erecta	Pointe	S. aureus (Gram +ve) E. coll (Gram -ve) S. typht (Gram -ve) S. flevneri (Gram -ve) S. aureus (Gram +ve)	***	- + + +	· · · ·		11:11:		
			tha (cham tve)	+++	+++	+++	+	+++		۲

Table 3: Antifungal activity of different solvent extracts of

Sr. No.	Plant Species	Plant Part	10	ent extracts of screened	braura at	gamai loui	Turigati a		.Ilesan	
		THE PARTY		Control (Flucanazole)	Ethanol	Methanol	Chloroform	Acetone		
			admid allernata	+++	++	++			<u> </u>	
١.	Vitex negundo	Leaves	Leaves	Penicillium notatum	+++	11	-	434	- + +	
		Aspergillus nicer	+++	++	- · ·	+				
			Penicillium digitatum	+++					++	
2. Nanthium st		umarium Seed	Alternaria alternata	+1-1	+	++	+++		++	
	Nanthium strumarium		Penicillium notatum	+1+	+	+	++		<del></del>	
•••	, , , , , , , , , , , , , , , , , , , ,		Aspergillus niger	+1+	•	·				
127.			Penicillium digitatum		+++	+		++		
		Seed	Alternaria alternata	+++	-	·	++	++-	++	
	Caesalpinia		Penicillium notatum	++-+	++	+			•	
3.	bonduc		Aspergillus niger	+++	•		111	<del></del>		
			Penicillium digitatum		+	+	++		-	
		Seed	Alternaria alternata	+++		<u> </u>		++	+	
	Mucuna		Penicillium notatum	++-+	+	+_	++			
4.	pruriens		Aspergillus niger	+- -+		<u> </u>			-	
	prurens		Penicillium digitatum	+++	-	•	++	+	ે.	
			Alternaria alternata	+++	++	+		+++	7-	
1	Tagetes	Petals	Penicillium notatum	+++	+++	+++	++	111	-	
5.				+++	· -	<u> </u>	<del>-</del> -	<del></del>	+	
••	erecta		Aspergillus niger Penicillium digitatum		++	+	+			

Acknowledgments

This work was financially supported by the Department of Science and Technology (DST), Government of India, New Delhi. The authors would like to thanks Principal, Nutan College, Selu, (MS), India and Authorities of Dr. Babasaheb Ambedkar Marathwada University, Aurangabad (MS), India for providing laboratory facility and National Chemical Laboratory (NCL), Pune (MS), India for supplying bacterial species as well as Department of Botany, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad (MS), India for supplying fungal strains,

### References

- Chávez-Quintal P. González-Flores T, Rodríguez-Buenfil I, Gallegos-Tintore S. Antifungal activity in ethanolic extracts of Carica papaya L. ev. Maradol leaves and seeds. Indian journal of microbiology. 2011; 51(1):54-60.
- 2. Chowdhury JA, Islam MS, Asifuzzaman SK, Islam MK. Antibacterial and cytotoxic activity screening of leaf extracts of Vitex negundo (Fam: Verbenaceae). Journal of Pharmaceutical Sciences and Research. 2009; 1(4):103-
- 3. Adwan G, Mhanna M. Synergistic effects of plant extracts and antibiotics on Staphylococcus aureus strains isolated from clinical specimens. Middle-East Journal of Scientific Research, 2008; 3(3):134-139.
- Djeussi DE, Noumedem JA, Seukep JA, Fankam AG, Voukeng IK, Tankeo SB et al. Antibacterial activities of selected edible plants extracts against multidrug-resistant Gram-negative bacteria. BMC Complementary and Alternative Medicine, 2013; 13(1):164.
- Rios JL, Recio MC. Medicinal plants and antimicrobial activity. Journal of Ethnopharmacology. 2005; 100(1, 2):80-84. ~ 1652 ^

- 6. Argal A, Pathak AK. CNS activity of Calatropis gigantea roots. Journal of ethnopharmacology. 2006;
- Khokra SL, Prakash O, Jain S, Aneja KR, Dhingra Y. Essential oil composition and antibacterial studies of Vitex negundo Linn. extracts. Indian Journal of Pharmaceutical Sciences, 2008; 70(4):522-526.
- Aswar PB, Khadabadi SS, Kuchekar BS, Rajurkar RM, Saboo SS, Javarkar RD. In-vitro evaluation of antibacterial and anti-fungal activity of Vitex nigundo Leaflets. Ethnobotanical (Verbenaceae).
- Panda SK, Thatoi HN, Dutta SK. Antibacterial activity and phytochemical screening of leaf and bark extracts of Vitex negundo L. from similipal biosphere reserve, Orissa. Journal of Medicinal Plants Research. 2009;
- 10. Gautam LM, Shrestha SL, Wagle P, Tamrakar BM. 3(4):294-300. Chemical constituents from Vitex negundo (Linn.) of nepalese origin. Scientific World. 2008; 6(6):27-32.
- 11. Nagarsekar KS, Nagarsenker MS, Kulkarni SR. Evaluation of composition and antimicrobial activity of supercritical fluid extract of leaves of Vitex negundo. Indian Journal of Pharmaceutical Sciences. 2010;
- 12. Sharma A, Tyagi S, Nag R, Chaturvedi A, Nag TN. Antimicrobial activity and cellular toxicity of flavonoid extracts from Pongamia pinnata and Vitex negundo. Romanian Biotechnological Letters. 2011; 16(4):6396-
- 13. Mahmud S, Sharcef H, Farrukh U, Kamil A, Rizwani GH. Antifungal activities of Vitex negundo Linn. Pakistan Journal of Botany. 2009; 41(4):1941-1943.

PRINCIPÁL Nutan Mahavidyalaya SELU, Dist. Parkhani



Sharma SR, Singh D, Khan FA, Swarankar CP, Bhagwan PS. Anthelmintic activity of Xanthium strumarium against Haemonchus contortus infection in sheep. Indian Journal of Animal Sciences. 2003; 73:342-4

Lavault M, Landreau A, Larcher G, Bouchara JP, Pagniez F, Le Pape P et al. Antileishmanial and activities of xanthanolides isolated from Xanthium macrocarpum. Fitoterapia. 2005; 76(3-

16. Favier LS, Maria AO, Wendel GH, Borkowski EJ, Giordano OS, Pelzer L et al. Anti-ulcerogenic activity of sesquiterpenes cavanillesii in rats. Journal of ethnopharmacology. 2005;

17. Kim IT, Park YM, Won JH, Jung HJ, Park HJ, Choi JW, et al. Methanol extract of Xanthium strumarium L. possesses anti-inflammatory and anti-nociceptive activities. Biological and Pharmaceutical Bulletin. 2005;

18. Kamboj A, Saluja AK. Phytopharmacological review of Xanthium strumarium L. (Cocklebur). International Journal of Green Pharmacy. 2010; 4(3):129-139.

Kim YS, Kim JS, Park SH, Choi SU, Lee CO, Kim SK, et al. Two cytotoxic sesquiterpene lactones from the leaves of Xanthium strumarium and their in vitro inhibitory activity on farnesyltransferase. Planta Medica. 2003; 69(4):375-377.

20. Srinivas PV, Rao RU, Venkateshwarulu EL, Kumar AC. Phytochemical screening and in vitro antimicrobial investigation of the methanolic extract of Xanthium strumarium leaf. International Journal of Drug Development and Research. 2011; 3:286-293.

21. Verma P, Verma A. Evaluation of antibacterial activity of different parts of Tagetes erecta. International Journal of Pharmacy and Life Sciences. 2012; 3(6):1766-1768.

- 22. Packianathan N, Karumbayaram S. Formulation and Evaluation of Herbal Hair Dye: An Ecofriendly Process. Journal of Pharmaceutical Sciences and Research, 2010; 2(10):648-656.
- 23. Cantrill R. Lutein from Tagetes erecta L. for use in foods for particular nutrition uses. The EFSA Journal. 2006; 315(1):1-12.
- Nikkon F, Habib MR, Karim MR, Ferdousi Z, Rahman MM, Haque ME. Insecticidal activity of flower of Tagetes erecta L. against Tribolium castaneum (Herbst). Research Journal of Agriculture and Biological Sciences. 2009; 5(5):748-753.
- 25. Sembiring EN, Elya B, Sauriasari R. Phytochemical screening, total flavonoid and total phenolic content and antioxidant activity of different parts of Caesalpinia bonduc (L.) Roxb. Pharmacognosy Journal. 2018; 10(1):123-127.
- 26. Navanit PS. Phytochemical extraction and antibacterial studies of Caesalpinia bonducella seed extracts. Mapana-Journal of Sciences. 2014; 13(4):47-54.
- 27. Nadkarni KM. Indian Materia Medica, 3rd ed. Vol. I, Bombay, Popular Prakashan; 1986, 335-337.
- 28. Kirtikar KR, Basu BD. Indian medicinal plants. 2nd ed. Dehradun: International book distributor; 1987, 763-7.
- Anonymous. The Wealth of India, Raw materials, Vol. III, CSIR, New Delhi; 1992, 179.
- 30. Sathiyanarayanan L, Arulmozhi S. Mucuna pruriens Comprehensive Review. Pharmacognosy Reviews. 2007; 1(1):157-162.

~ 1653 ~

31. Lampariello LR, Cortelazzo A, Guerranti R, Sticozzi C, Valacchi G. The magic velvet bean of Mucuna pruriens. Journal of Traditional and Complementary Medicine.

32. Harborne AJ. Phytochemical Methods- A guide to modern techniques of plant analysis 3rd edn. Chapman and Hall, Springer Science & Business Media, New York. 1998, 1-150.

33. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic Susceptibility Testing by a Standardized Single Disk Method. American Journal of Clinical Pathology. 1966;

34. Noumedem JA, Mihasan M, Lacmata ST, Stefan M, Kuiate JR, Kuete V. Antibacterial activities of the methanol extracts of ten Cameroonian vegetables against bacteria. Gram-negative multidrug-resistant Medicine. and Alternative Complementary

13(1):26. antimicrobial Plant products as agents. Clinical microbiology reviews. 1999; 12(4):564-MM. 35. Cowan

36. Rose CM, Cathrine L. Preliminary phytochemical screening and antibacterial activity on Vitex negundo. International Journal of Current Pharmaceutical Research. 2011; 3(2):99-101.

37. Lolge SC, Zanke SP, Patil DR, Zambare SP. In vitro Antimicrobial Activity and Phytochemical Screening of Local Plants. Journal of Pharmacy and Pharmacology. 2016; 4:151-154.

38. Islam S, Akhtar M, Parvez S, Alam J, Alam FM. Antitumor and antibacterial activity of a crude methanol leaf extract of Vitex negundo L. Archives of Biological Sciences. 2013; 65(1):229-238.

39. Devkota A, Das RK. Antibacterial Activities of Xanthium strumarium L. Journal of Natural History Museum. 2015; 29:70-77.

40. Malik F, Hussain S, Mirza T, Hameed A, Ahmad S, Riaz H et al. Screening for antimicrobial activity of thirtythree medicinal plants used in the traditional system of medicine in Pakistan. Journal of Medicinal Plants Research. 2011; 5(14):3052-3060.

41. Khuda F, Iqbal Z, Khan A, Nasir F, Khan MS. Validation of some of the ethnopharmacological uses of Xanthium strumarium and Duchesnea indica. Pakistan Journal of Botany. 2012; 44(4):1999-1201

> PRINCIPAL. Nutan Mahavidyalaya SELU, Dist. Parbhani

